

Central European hardwood trees in a high-CO₂ future: synthesis of an 8-year forest canopy CO₂ enrichment project

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Summary

1. Rapidly increasing atmospheric CO₂ is not only changing the climate system but may also affect the biosphere directly through stimulation of plant growth and ecosystem carbon and nutrient cycling. Although forest ecosystems play a critical role in the global carbon cycle, experimental information on forest responses to rising CO₂ is scarce, due to the sheer size of trees.

2. Here, we present a synthesis of the only study world-wide where a diverse set of mature broadleaved trees growing in a natural forest has been exposed to future atmospheric CO₂ levels (*c.* 550 ppm) by free-air CO₂ enrichment (FACE). We show that litter production, leaf traits and radial growth across the studied hardwood species remained unaffected by elevated CO₂ over 8 years.

3. CO₂ enrichment reduced tree water consumption resulting in detectable soil moisture savings. Soil air CO₂ and dissolved inorganic carbon both increased suggesting enhanced below-ground activity. Carbon release to the rhizosphere and/or higher soil moisture primed nitrification and nitrate leaching under elevated CO₂; however, the export of dissolved organic carbon remained unaltered.

4. Synthesis. Our findings provide no evidence for carbon-limitation in five central European hardwood trees at current ambient CO₂ concentrations. The results of this long-term study challenge the idea of a universal CO₂ fertilization effect on forests, as commonly assumed in climate–carbon cycle models.

Key-words: CO₂ fertilization, coupled climate–carbon cycle model, ecosystem carbon cycling, elevated CO₂, free-air CO₂ enrichment (FACE), global carbon cycle

Introduction

At present, the Earth's vegetation is exposed to atmospheric CO₂ concentrations that have been unprecedented over the past 15 million years (Tripathi, Roberts & Eagle 2009). By default, land ecosystems are probably absorbing *c.* 25–30% of the annual anthropogenic CO₂ emissions, which have dramatically increased over the last decade and are now tracking the worst-case IPCC scenario (A1F1, Meehl *et al.* 2007; LeQuéré *et al.* 2009). Forests are major potential contributors to this terrestrial carbon (C) sink as they account for nearly 90% of

the terrestrial biomass and about half of terrestrial net primary productivity (NPP; Roy, Saugier & Mooney 2001). Including soils, forest ecosystems accommodate roughly twice the amount of C held in the atmosphere (Jobbágy & Jackson 2000; Roy, Saugier & Mooney 2001). Given their prominent role in the global C cycle, the responses of trees and forests to rising atmospheric CO₂ are essential to understand how C cycling will change in a future high-CO₂ world and how these changes might feedback to other ecosystem processes and the climate system. Future climate predictions are highly sensitive to changes in the global C cycle in response to increasing atmospheric CO₂ (Friedlingstein *et al.* 2006). Coupled climate–carbon models commonly assume a CO₂ fer-

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tilization effect on plant growth that is further assumed to eventually result in enhanced C storage in organic matter, a step that lacks a theoretical as well as an empirical foundation (a flux vs. pool confusion, see Körner 2006). Any increase in terrestrial C storage would reduce the rate of CO₂ accumulation in the Earth's atmosphere and thus potentially mitigate the trajectory of climate change. However, the magnitude of the CO₂ fertilization effect and its continuity into the future are a matter of debate (Matthews 2007). There is plenty of evidence that elevated atmospheric CO₂ can stimulate plant growth when nutrients and other resources are not limiting, as in many agricultural and horticultural environments. Growth responses to CO₂ enrichment in near-natural systems, where growth is often water- or nutrient-limited, vary widely but are certainly smaller than anticipated and tend to decrease over time, especially in forests (Leuzinger *et al.* 2011; Norby & Zak 2011). Even under favourable soil conditions, growth in elevated CO₂ may eventually lead to progressive nitrogen (N) limitation through accelerated fixation of N in organic matter and thus cause downregulation of forest growth responses (Luo *et al.* 2004; Norby & Zak 2011). Such interactions of elevated CO₂ with biogeochemical processes that operate on longer time scales cannot be revealed in short-term studies in decoupled systems and highlight the importance of long-term CO₂ enrichment experiments in natural forest settings. CO₂-induced growth stimulation may only translate into enhanced forest biomass C sequestration if trees grow larger by maturing more rapidly and also show a prolongation of their life span (Körner 2009). However, any CO₂ fertilization effect may be offset by effects on tree longevity (Bugmann & Bigler 2011), because carbon storage in forest biomass is, in essence, an issue of tree demography that cannot be explored in CO₂ enrichment experiments.

Owing to the size of trees, forest data are critically under-represented in the experimental CO₂ research literature, with most evidence originating from young trees and premature stands. Further, tree size restricts the tree species diversity that can be accommodated in a given test plot, which conflicts with the well-known species specificity of tree CO₂ responses (Keel *et al.* 2007; Leuzinger & Körner 2007; Dawes *et al.* 2011; Smith *et al.* 2013a). Here, we present a synthesis of the effects of 8 years of CO₂ enrichment in the so far only natural, diverse forest tested, including novel, previously unpublished data on tree-ring increments, stable C isotope signatures in wood, dissolved C fluxes in soil water and N availability (foliar δ¹⁵N). In addition, we extended the published 2001–2004 time series for leaf litter production, leaf traits and soil CO₂ dynamics (Körner *et al.* 2005; Keel, Siegwolf & Körner 2006) with more recent data from 2005–2008 and report the results of the new multiyear analyses.

This long-term free-air CO₂ enrichment (FACE) experiment was established in north-west Switzerland to test whether rising atmospheric CO₂ concentrations will stimulate tree growth and the forest C cycle and alter N availability. With the aid of a construction crane, we installed a fine web of perforated tubes releasing pure CO₂ into the crowns of 11 35-m-tall deciduous trees to simulate a future atmosphere containing

c. 550 ppm CO₂ (Pepin & Körner 2002). The study comprised *Fagus sylvatica* and *Quercus petraea* – two dominant forest trees in Central Europe – as well as the subdominant *Carpinus betulus*, *Tilia platyphyllos* and *Acer campestre*.

Materials and methods

STUDY SITE

The Swiss Canopy Crane (SCC) FACE site was established in 1999 in an c. 100-year-old mixed forest, situated 15 km south of Basel, Switzerland (47°28'N, 7°30'E, 550 m.a.s.l.). The closed-canopy stand is about 30–35 m tall and has a tree density of 415 trees ha⁻¹ (breast height diameter ≥ 0.1 m), a peak leaf area index of 5 (without counting the largely herbaceous forest understorey) and a basal area of 46 m² ha⁻¹ (Pepin & Körner 2002). *Fagus sylvatica* L. (European beech) and *Quercus petraea* (Matt.) Liebl. (sessile oak) dominate the forest canopy, accompanied by subdominant tree species such as *Carpinus betulus* L. (hornbeam), *Tilia platyphyllos* Scop. (large-leaf linden), *Acer campestre* L. (field maple), *Prunus avium* L. (cherry) and four species of conifers [*Picea abies* (L.) Karst., *Larix decidua* Mill., *Pinus sylvestris* L., *Abies alba* Mill.]. With the help of a 45-m-tall construction crane, 12 deciduous trees were initially subjected to canopy CO₂ enrichment, beginning in autumn 2000 (three *F. sylvatica*, three *Q. petraea*, three *C. betulus*, one *T. platyphyllos*, one *A. campestre* and one *P. avium*). Due to its near to off-site position under the crane's reach and some storm damage, the single *P. avium* tree had to be excluded from the CO₂ treatment in 2006, and a maple control tree was felled by a winter storm. The crowns of the study trees were exposed to an elevated concentration of atmospheric CO₂ (c. 550 ppm) during daylight hours throughout the growing season using a canopy FACE system (Pepin & Körner 2002). Intentionally, the understorey vegetation did not receive CO₂ enrichment, and canopy CO₂ enrichment 25–35 m above-ground did not leak to the ground, hence offering unbiased reference conditions for obtaining soil ¹³C isotope signals (Keel, Siegwolf & Körner 2006). A larger number of control trees were selected at sufficient distance to the CO₂-enriched area. The applied CO₂ was of fossil origin and thus depleted in ¹³C (−29.33 ± 0.23‰, 8-year mean ± SE, i.e. delivered at a constant quality over the study years), which allowed us to trace the fate of the newly assimilated C. The experiment was terminated at the end of the 2008 growing season.

LEAF LITTER PRODUCTION, LEAF TRAITS AND FOLIAR ¹⁵N ANALYSIS

Leaf litter was collected from 44 litter traps (0.5 m²) arranged in a 6-m grid within the study area. In autumn, traps were emptied biweekly, and the litter was oven-dried at 80 °C for at least 48 h and then weighed. Litter fall in the year 2001 was considered to still represent foliage produced under pre-treatment conditions. Hence, litter fall of 2001 was averaged with data from 1999 and 2000 to account for natural spatial variability of litter fall over 3 years before the start of CO₂ enrichment.

In summer (June–July), 10 leaf discs (1.2 cm²) per tree were collected from the crane gondola for determination of specific leaf area (SLA), non-structural carbohydrates (NSC), nitrogen concentrations and the ¹⁵N/¹⁴N isotope ratio (δ¹⁵N) in the top of the canopy. Leaf discs were dried at 80 °C for at least 48 h and then weighed for biomass quantification. After grinding, all samples from one tree were

merged for leaf N analysis using a CHN-analyser (Vario EL III, Elementar Analysensysteme GmbH, Hanau, Germany) and for NSC analysis based on an enzymatic starch digestion followed by a spectrophotometric glucose test after invertase and isomerase addition. Foliar $\delta^{15}\text{N}$ was analysed with an isotope ratio mass spectrometer (Delta S, Thermo Finnigan Mat), which was linked via a variable open split interface (CONFLO II) to an elemental analyser (EA 1108, Carlo Erba, Milano, Italy). Natural ^{15}N abundance was expressed in the δ -notation: $\delta^{15}\text{N} = R_{\text{sample}}/R_{\text{standard}} - 1$, where R is the molar ratio of ^{15}N to ^{14}N for the sample and the standard, respectively.

RADIAL INCREMENT AND TREE-RING $\delta^{13}\text{C}$

We sampled 2–3 wood cores per tree using a stainless steel wood corer to assess radial increment and $\delta^{13}\text{C}$ in growth rings. Tree-ring widths were measured using an automated bench with a stereomicroscope (Lintab/TSAP, Rinntech, Heidelberg, Germany). Dendrochronological analyses were performed using the dplR library (Bunn 2008) within the R environment (R 2.12.0, R Development Core Team 2011). From the raw tree-ring series, we created a mean value chronology for each of the sampled tree species by averaging each year's radial increment using Tukey's biweight robust mean. One of the advantages of working with mature trees is that their individual pre-treatment history of growth is conserved in the tree-ring chronology. This is of particular significance in a case, where large spatial replication finds practical limitation (would require several crane sites). We used pre-treatment growth of each individual to standardize its growth rate during the 8 years of experimentation. However, we also show the non-standardized data, illustrating that the results are robust, irrespective of standardization. Wood samples of yearly growth rings for $\delta^{13}\text{C}$ analysis were collected from wood cores with a scalpel under a stereomicroscope. Subsequently, the sample material was oven-dried at 80 °C for 48 h and then ground in a steel ball mill (Retsch MM 2000, Haan, Germany). 0.6–0.8 mg of the dried powder was filled into tin capsules and then combusted in an elemental analyser (EA-1110, Carlo Erba Thermoquest, Milan, Italy). Continuous-flow isotope ratio mass spectrometry was employed for ^{13}C isotopic analysis as described above. The precision of $\delta^{13}\text{C}$ analyses was < 0.1‰. The isotope values are expressed in the δ -notation: $\delta^{13}\text{C} = R_{\text{sample}}/R_{\text{standard}} - 1$, where R is the molar ratio of ^{13}C to ^{12}C for the sample and standard (VDBD), respectively.

VOLUMETRIC SOIL WATER CONTENT AND SOIL WATER ANALYSES

Soil water content at 0–10 cm depth was logged at 6-hourly intervals using 'ECH₂O Probes' (EC-10, Decagon Devices Ltd., Pullman, Washington, DC; ambient CO₂: $n = 20$, elevated CO₂: $n = 15$). Individual sensor readings were standardized to their maximum value.

Ceramic suction cups were installed along three transects through the experimental area, each running under individuals of each of the main tree species, *F. sylvatica*, *Q. petraea* and *C. betulus* (Schleppi *et al.* 2012). At 10–12 locations along each transect, two ceramic cups were installed at a depth of 15 cm and 1 or 2 at 5 cm. All suction cups were connected via a sampling bottle to vacuum systems reaching approximately –400 hPa. A soil solution sample from each location and depth was collected on average every 3 weeks, except when temperatures were below freezing point in winter. The samples were immediately refrigerated, then filtered (0.45 μm) and analysed for nitrate (NO₃⁻), ammonium (NH₄⁺), dissolved organic nitrogen (DON), organic carbon (DOC) and inorganic C (DIC). At 17 different

dates during the experiment, the abundance of ^{13}C in DIC was measured by releasing the CO₂ through acidification with phosphoric acid (H₃PO₄) in gas-tight vials (see Schleppi *et al.* 2012 for details). During summer, ion-exchange resin bags were buried near the suction cups to provide integrated measurements of available N. After 6 months in the soil, they were retrieved and analysed for NH₄⁺ as well as for NO₃⁻ and ^{15}N abundance (Schleppi *et al.* 2012).

Vertical fluxes of C and N below the litter layer and at depths of 5 and 20 (15) cm were estimated by multiplying the DOC, DON and NO₃⁻ concentrations with water fluxes obtained with the COUP simulation model (Jansson & Karlberg 2004). The model was parameterized using the organic C content, density and texture of different soil layers, as well as the height and leaf area index of the forest stand. The input variables were air temperature, precipitation, vapour pressure deficit, wind speed and net radiation.

STABLE CARBON ISOTOPE ANALYSIS IN SOIL AIR CO₂

The stable carbon isotope ratio ($\delta^{13}\text{C}$) and CO₂ concentration in soil air were monitored monthly during growing seasons using a grid of 86 PVC gas wells whose tops were sealed with a silicone rubber septum ($L \times D = 12 \times 2$ cm; 3×3 m grid spacing in the FACE area, 6×6 m in the control area). Soil air $\delta^{13}\text{C}$ analyses were performed using a mass spectrometer (Delta Plus XL) linked to a Gasbench-II periphery (both Thermo Finnigan, Bremen, Germany). Soil air CO₂ concentration was estimated from instrument calibration curves based on CO₂ reference gases (340 and 4995 ppm). For each sampling date and CO₂ treatment, the Keeling plot approach was applied to ascertain the $\delta^{13}\text{C}$ of soil air CO₂, that is, we applied linear regression models using measured $\delta^{13}\text{C}$ values as response and the inverse soil air CO₂ concentrations as explanatory variable. The intercept of these models corresponded to the mean soil air $\delta^{13}\text{C}$ and the standard error of the intercept served as a measure of spread. Isotope fractionation due to slower diffusion of the heavier $^{13}\text{CO}_2$ was accounted for by subtracting 4.4‰ (cf. Steinmann *et al.* 2004; Keel, Siegwolf & Körner 2006).

STATISTICAL ANALYSIS

All statistical analyses and graphics were performed using R version 2.12.0 (R Development Core Team 2011). Linear mixed effects models (package *nlme*) fitted by restricted maximum likelihood (REML) were applied to assess the effects of elevated atmospheric CO₂ on radial increment, litter fall, leaf traits, foliar $\delta^{15}\text{N}$, soil air $\delta^{13}\text{C}$ and soil air CO₂ concentration. These models contained the factors 'species', 'CO₂', 'year' and their interactions as fixed term and the factor 'month' nested within 'year' or simply 'tree' (in the case of radial increment and foliar $\delta^{15}\text{N}$) as repeated measure effects (random term). Homogeneity of variance was examined by inspection of the residuals after model fitting (plots of standardized residuals vs. fitted values and explanatory variables). Heterogeneous within-group variances were modelled either using a constant variance function (varIdent function, package *nlme*), a power variance function (varPower, package *nlme*), an exponential variance function (varExp, package *nlme*) or a combination thereof. Quantile–quantile plots were used to test for normally distributed model residuals and random effects. An autocorrelation function was used to check model residuals for temporal autocorrelation, and an autoregressive moving average (ARMA) correlation structure was employed to model dependence among observations. Backwards model selection to determine the optimal fixed term was based on comparing nested models using the Akaike Information

Criterion (AIC) and likelihood ratio tests. For the analysis of tree-ring $\delta^{13}\text{C}$ in CO_2 -enriched trees, we applied a generalized additive mixed model (GAMM) treating 'tree individual' as random effect to account for repeated measures. Temporal autocorrelation was modelled using a first-order autoregressive structure (AR1), and variance heterogeneity was modelled using an exponential variance function (varExp). The standardized soil moisture data were also analysed using GAMMs. The specific sensor locations within the control and CO_2 -enriched area were modelled as a random effect, and temporal autocorrelation was modelled using an autoregressive model of order 1 (AR-1 autocorrelation structure). The significance of the effect of CO_2 enrichment was tested using a likelihood ratio test comparing two nested GAMMs; a first model with a common smoother term for ambient and elevated CO_2 vs. a second model employing individual smoothers for each treatment.

The local effect of the CO_2 treatment in the soil around suction cups and resin bags was quantified by the seasonal depletion of their DIC in ^{13}C (Schleppi *et al.* 2012). Concentrations in the soil solution were then interpreted based on a generalized linear model including this ^{13}C -derived index and the duration of the treatment. The interaction between both was tested statistically to show whether the effect of soil locations changed over time as related to the CO_2 enrichment. Due to their very high variability, NO_3^- concentrations in the soil solution were log-transformed prior to analysis, which yielded homoscedastic residuals with a distribution not significantly departing from normality.

Results

LEAF LITTER PRODUCTION AND LEAF TRAITS

Annual leaf litter fall collected in traps did not change in response to canopy CO_2 enrichment (Fig. 1). Foliar non-structural carbohydrates (NSC) in *F. sylvatica*, *C. betulus* or any of the two unreplicated species (*T. platyphyllos*, *A. campestre*) were unaffected by elevated atmospheric CO_2 . In CO_2 -enriched *Q. petraea* leaves however, NSC accumulated significantly and consistently throughout the eight experimental years (on average +22%, $\text{CO}_2 \times \text{species}$, $P < 0.0001$), a signal that disappeared in the first post-treatment year. Specific leaf area in the upper canopy showed significant

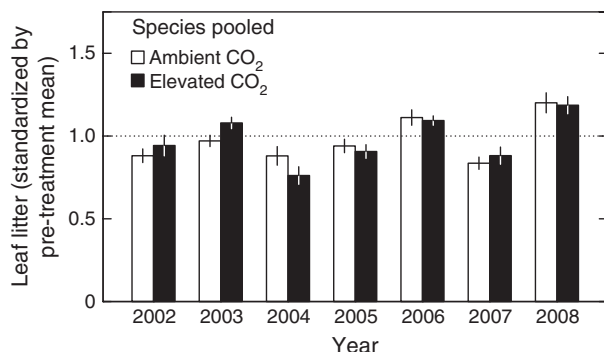


Fig. 1. Leaf litter production of CO_2 -enriched and control trees (standardized by the 3-year-pre-treatment mean for each 1 m^2 litter trap 1999–2001). Means \pm SE, ambient CO_2 $n = 32$, elevated CO_2 $n = 12$.

interspecific variation ($P < 0.001$) but was not consistently affected by elevated CO_2 (Fig. 2). Leaf N in the replicated *F. sylvatica*, *Q. petraea* and *C. betulus* trees remained unaffected by elevated CO_2 (Fig. 2). In the second half of the CO_2 -enrichment period (2005–2008), the two unreplicated species, *A. campestre* and *T. platyphyllos*, showed reductions in leaf N concentration under elevated CO_2 (on a NSC-free dry matter basis, species $\times \text{CO}_2$, $P < 0.001$), but already in the first post-treatment year, foliar N had returned to control levels.

RADIAL INCREMENT AND TREE-RING $\delta^{13}\text{C}$

CO_2 enrichment had no significant effect on radial increment growth (expressed as raw tree-ring width and standardized by the 10-yr pre-treatment mean) in the dominant *F. sylvatica* and *Q. petraea* trees and the subdominant *Tilia* tree ($P = 0.52$, Fig. 3). However, not surprisingly, in some study years, there was significant interspecific variation in radial growth (species \times year interaction, $P < 0.01$). Tree-ring series obtained from *C. betulus* and *A. campestre* were not reliably dateable and therefore had to be excluded from the analysis. The incorporation of recently assimilated (isotopically labelled) C into newly formed wood occurred remarkably fast in our study trees (Fig. 4). Already after the first year of canopy enrichment with ^{13}C -depleted CO_2 , the stable C isotope ratio in growth rings of CO_2 -treated trees had dropped by c. 4‰ and remained at a significantly lower level throughout the experimental period ($P < 0.001$, Fig. 4). In the first tree-ring produced after the termination of CO_2 enrichment (i.e. cessation of stable carbon isotope labelling), the $\delta^{13}\text{C}$ signal had already become less negative, and after the third post-treatment year, the tree-ring $\delta^{13}\text{C}$ signature had returned to pre-treatment values (Fig. 4).

SOIL WATER SAVINGS

During growing seasons, volumetric water content was significantly higher in soil under trees receiving CO_2 enrichment compared to soil surrounding control trees resulting from reduced transpiration under elevated CO_2 (Fig. 5, $L = 16.74$, $P < 0.001$, $\Delta\text{AIC} = 12.75$). For example, in 2007, before the spring leaf flush, soil moisture was similar under all trees regardless of treatment. However, with advancing leaf maturation and thus rising canopy transpiration, soil water content became progressively lower around control trees compared to trees exposed to elevated CO_2 . Replenishment of soil water resources by a number of consecutive rainy days or heavy rainfall events transiently offset treatment-related differences in soil moisture (Fig. 5). After leaf fall, soil water content under control trees and CO_2 -enriched trees gradually converged again.

SOIL WATER CHEMISTRY

The depletion of ^{13}C in the dissolved inorganic carbon sampled in suction cups was on average $2.2 \pm 1.4\text{‰}$, ranging

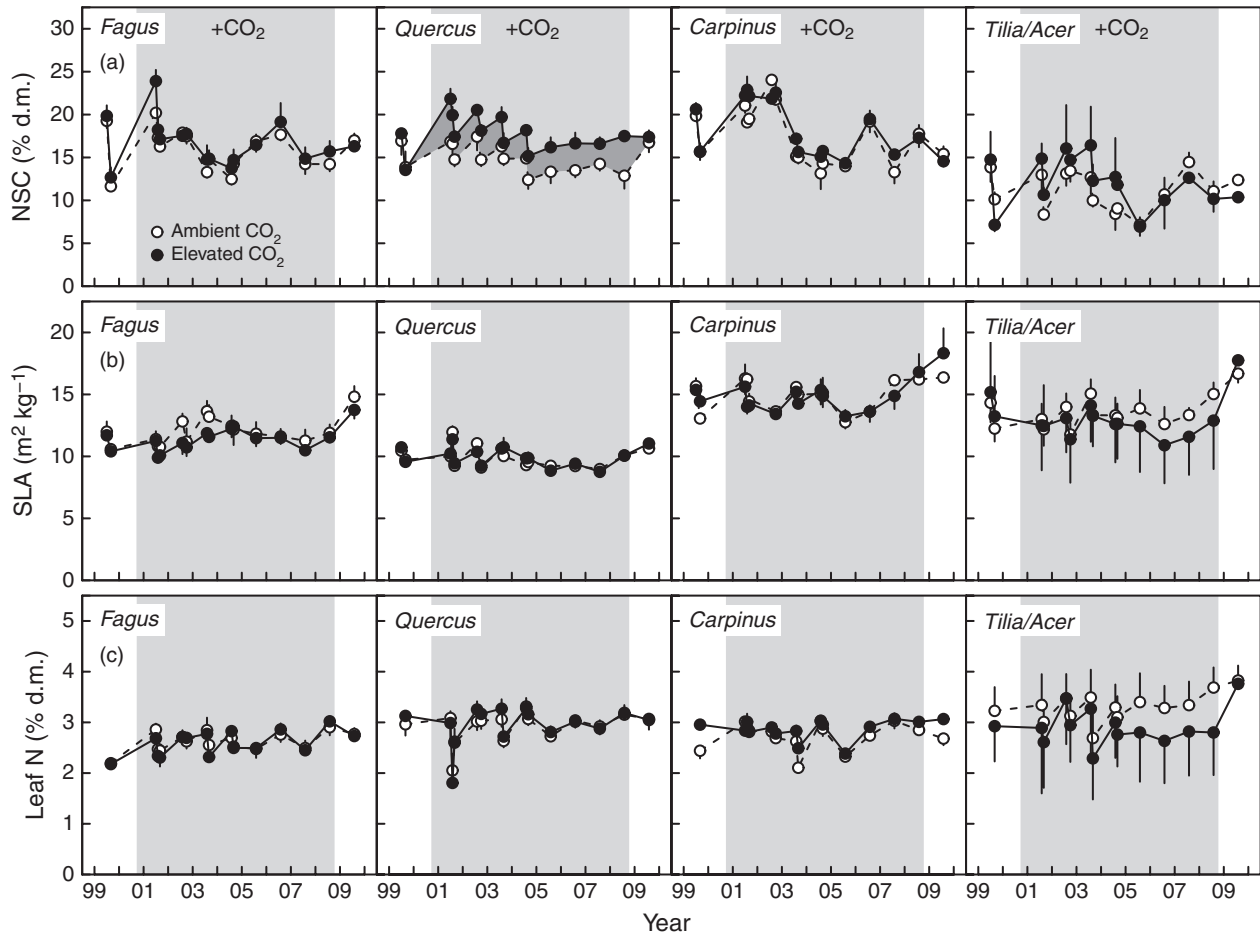


Fig. 2. Traits of fully expanded upper-canopy leaves produced under ambient (white) and elevated (black) atmospheric CO₂. Non-structural carbohydrate (NSC) content (a), specific leaf area (SLA) (b) and leaf nitrogen (N) concentration (c). SLA and leaf N are given on an NSC-free basis. Means \pm SE, ambient CO₂ $n = 3-5$, elevated CO₂ $n = 3$. For the single *Tilia platyphyllos* and *Acer campestre* trees, the pooled response is shown ($n = 2$ trees). The grey-shaded area indicates the CO₂-enrichment period. The dark grey area highlights the pronounced NSC response in CO₂-enriched *Quercus petraea* trees.

from 4.9 to 1.3‰. These values were used as an indicator of the degree of local exposure of soil pockets to the effects of canopy CO₂ enrichment across the treatment area. Their interaction with the duration of the experiment had a positive effect on nitrate concentrations ($P < 0.01$; Schleppei *et al.* 2012). Multiplying the measured concentrations at 15 cm depth by drainage water fluxes obtained with the COUP model resulted in an annual NO₃⁻-N leaching of 2.3 kg ha⁻¹ at control locations. At corresponding locations along the soil transects, the CO₂ enrichment increased this leaching by an additional 1.2 kg ha⁻¹. Amounts of nitrate captured by the resin bags confirmed this trend, showing a clear increase in free nitrate due to the CO₂ enrichment ($P < 0.0001$). In parallel, the natural abundance of ¹⁵N in this nitrate strongly increased (Schleppei *et al.* 2012).

In the topsoil at 5 cm depth, concentrations of DIC were increased by 50% under elevated CO₂ (time \times CO₂, $P = 0.0012$), while DOC concentrations declined by 20% during the eight treatment years (time \times CO₂, $P < 0.0001$). The corresponding effects on the fluxes of dissolved C cancelled each other, with CO₂ enrichment increasing fluxes of

DIC by 2.8 g C m⁻² a⁻¹, but decreasing DOC fluxes by 2 g C m⁻² a⁻¹ (Fig. 6). At 15-cm-soil depth, concentrations and fluxes of DIC and DOC were not affected by elevated CO₂; hence, the signal disappeared within the soil profile.

SOIL AIR $\delta^{13}\text{C}$ AND CO₂ CONCENTRATION

Throughout the eight-year study period, respiratory air in the soil pore space under CO₂-enriched trees remained isotopically labelled, also during the dormant seasons, when CO₂ enrichment was discontinued (treatment-related difference in $\delta^{13}\text{C}$ including the dormant seasons: 1.7 ± 0.12 ‰, 8-yr mean \pm SE, Fig. 7a), with the remainder of CO₂ release apparently resulting from older (pre-treatment) carbon. Relating this isotopic signature to the isometer reference $\delta^{13}\text{C}$ (obtained from C₄ grasses installed in the tree canopies) suggested that soil respiratory CO₂ under CO₂-treated trees consisted on average of *c.* 30% labelled (novel) C; however, the proportion of this recent C varied significantly between study years (CO₂ \times year interaction, $P < 0.01$).

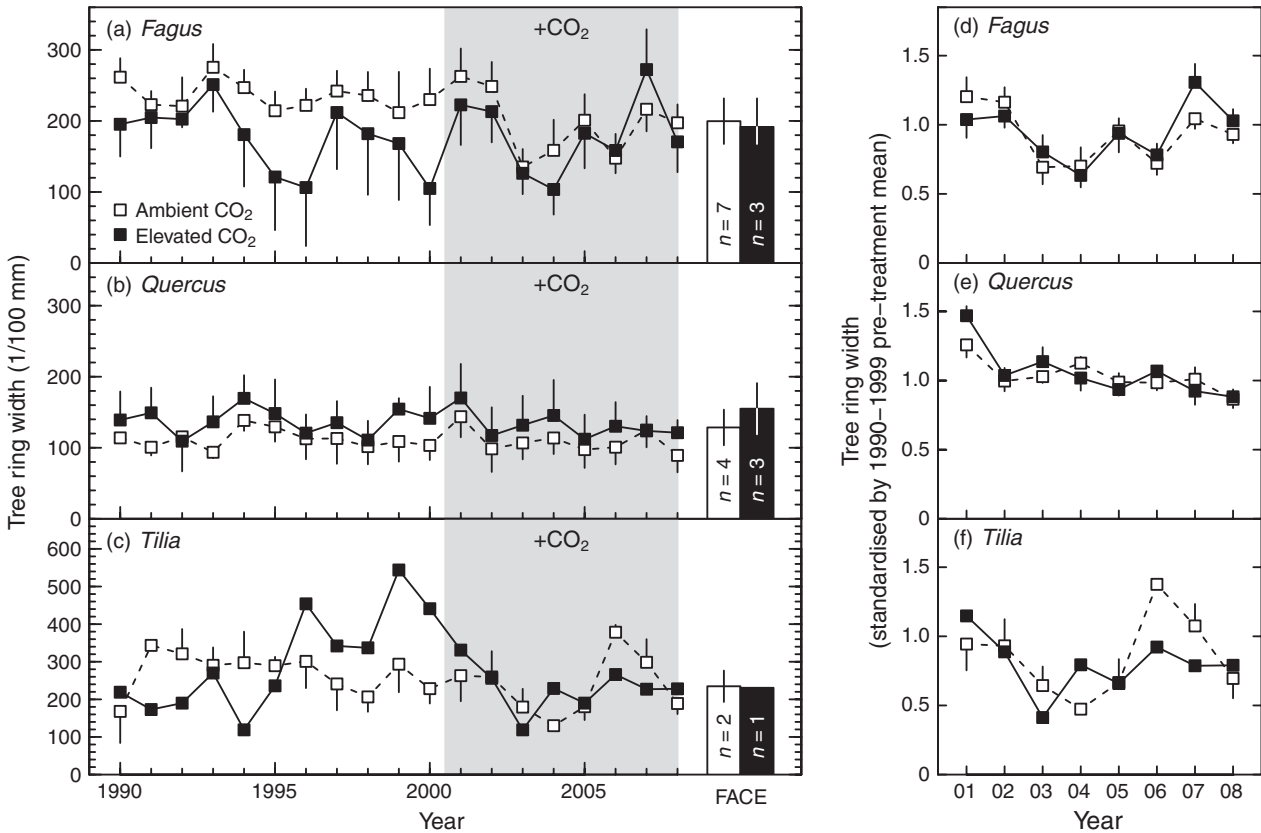


Fig. 3. Annual radial stem growth given as raw ring width (left panels) and standardized ring width (right panels, divided by the 1990–1999 pre-treatment mean) of dominant *Fagus sylvatica* (a, d), *Quercus petraea* (b, e) and subdominant *Tilia platyphyllos* trees (c, f) growing under ambient (dashed line, white symbols) and elevated (solid line, black symbols) atmospheric CO₂ in a closed-canopy temperate forest in northern Switzerland (ring widths of the remaining tree species were not dateable). The grey-shaded area indicates the CO₂-enrichment period (8 years). Bar chart inserts give the average ring width increment during the free-air CO₂-enrichment period (FACE).

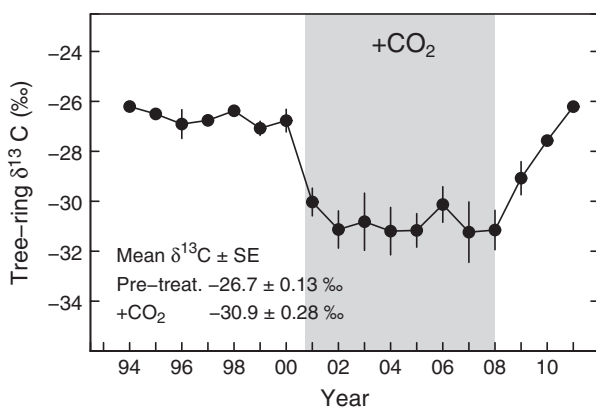


Fig. 4. Tree-ring $\delta^{13}\text{C}$ signals derived from CO₂-enriched *Fagus sylvatica*, *Quercus petraea* and *Tilia platyphyllos* trees. Means \pm SE, $n = 1\text{--}3$.

During growing seasons, CO₂ accumulated more strongly in the soil pore space under CO₂-enriched trees compared to controls, on average by 35% ($P < 0.001$, Fig. 7b). When sampling had been extended into the dormant season (in the years 2001–2003), treatment-related differences in soil air CO₂ concentrations disappeared and values were similar under all trees (Fig. 7b).

FOLIAR $\delta^{15}\text{N}$

We monitored foliar ^{15}N natural abundance (expressed as $\delta^{15}\text{N}$) in archived leaf samples of our tall study trees as a proxy for ecosystem N availability. Prior to the start of CO₂ enrichment, leaf $\delta^{15}\text{N}$ signatures were similar among all trees (Fig. 8). Throughout the entire CO₂-enrichment period, foliar $\delta^{15}\text{N}$ remained extraordinarily steady under elevated CO₂, while the $\delta^{15}\text{N}$ values of control foliage showed more variation and were overall slightly but significantly more negative (2001–2008 average: elevated CO₂ $-2.35 \pm 0.15\text{‰}$, ambient CO₂ $-2.82 \pm 0.18\text{‰}$, $P < 0.01$, Fig. 8). Of particular note was a pronounced drop in the $\delta^{15}\text{N}$ signal by roughly 0.7‰ in leaves of control trees in 2004, following a centennial heat wave in 2003.

Discussion

Combining novel data that became available at or after the end of canopy CO₂ enrichment, with data that had been collected during the entire 8-year experiment, permitted us to consolidate an overall message of this unique experiment. Using pre-treatment and post-treatment signals, those of ^{13}C in particular, illuminates the C dynamics in such tall trees.

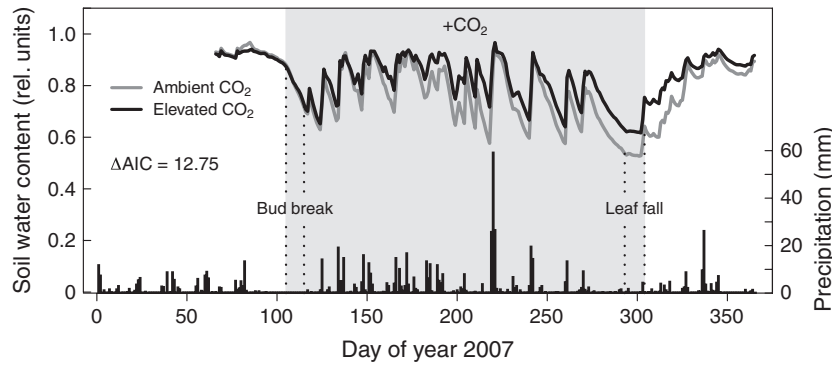


Fig. 5. Precipitation and volumetric soil water content under CO₂-enriched and control trees in 2007. The grey-shaded area designates the CO₂-enrichment period, and the dotted lines indicate the bud break and leaf fall period, respectively. The Δ AIC value is derived from a model comparison between a generalised additive mixed model (GAMM) with a common smoother term and a GAMM with individual smoother terms for ambient and elevated CO₂ (the large Δ AIC value lends strong support to the model with individual smoothers suggesting treatment-related differences, see Materials and Methods for details).

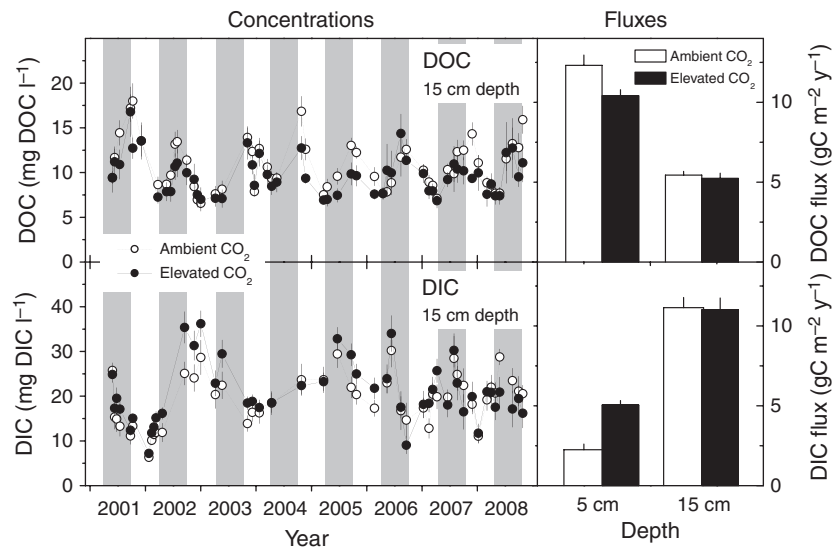
LITTER PRODUCTION AND LEAF-LEVEL C UPTAKE UNDER CO₂ ENRICHMENT

Based on leaf duration data (Asshoff, Zotz & Körner 2006) and annual leaf litter production, which both remained unaffected by high CO₂, we can conclude that canopy CO₂ enrichment did not alter the steady leaf area index of five in this mature forest, similar to what has been reported for a deciduous plantation forest in Tennessee (Oak Ridge FACE; Norby & Zak 2011). The strong sugar signal observed in *Q. petraea* foliage did not feedback negatively on photosynthetic performance, which is also in line with the findings for *Liquidambar styraciflua* at the Oak Ridge FACE site (Sholtis *et al.* 2004). In the eighth and final year of CO₂ enrichment, photosynthetic C uptake in the upper canopy was on average 42–48% higher in CO₂-enriched trees and showed no signs of downregulation (Bader, Siegwolf & Körner 2010), consistent with earlier findings from this and other forest FACE sites (Zotz, Pepin & Körner 2005; Liberloo *et al.* 2007; Darbah *et al.* 2010; Norby *et al.* 2010; Ellsworth *et al.* 2012). However, at the Oak Ridge FACE site, progressive nitrogen limitation has eventually caused a complete loss of the photosynthetic response to elevated CO₂ (Norby *et al.* 2010).

RADIAL GROWTH AND STABLE C ISOTOPE SIGNATURES IN TREE-RINGS UNDER ELEVATED CO₂

Elevated atmospheric CO₂ has often been reported to stimulate radial stem growth in young trees, at least temporarily, but it has also become evident that potential growth responses are highly species-specific and contingent on tree age, stand demography and the availability of resources other than CO₂ (Körner 2006; Norby & Zak 2011; Smith *et al.* 2013a). The forest soil at our SCC FACE site is nutrient-rich and according to accepted forestry standards the currently *c.* 110-year-old stand grows vigorously, yet our study trees showed no persistent growth stimulation in response to CO₂ enrichment, suggesting alternative pathways for the extra C assimilated under elevated CO₂. By contrast, *Pinus taeda* growing in a temperate plantation forest (Duke FACE) and young poplar saplings in bio-energy plantations on fertile ground (POP-FACE) showed sustained enhancement of radial stem growth under CO₂ enrichment (pine: 13–27% across study years, poplar: 20–29%, Moore *et al.* 2006; Liberloo *et al.* 2006). However, at the Oak Ridge FACE site, the initial stimulation in radial stem growth of *Liquidambar styraciflua* under elevated CO₂ rapidly shifted below-ground to increased fine root

Fig. 6. Inter-annual and intra-annual variation in dissolved organic carbon (DOC) and dissolved inorganic carbon (DIC) concentrations in the upper soil horizon (at 15 cm depth) under CO₂-enriched and control trees (left panels). Corresponding DOC and DIC fluxes at 5 and 15 cm depth (right panels). The grey-shaded rectangles indicate the CO₂-enrichment periods that correspond to the 6-month growing seasons.



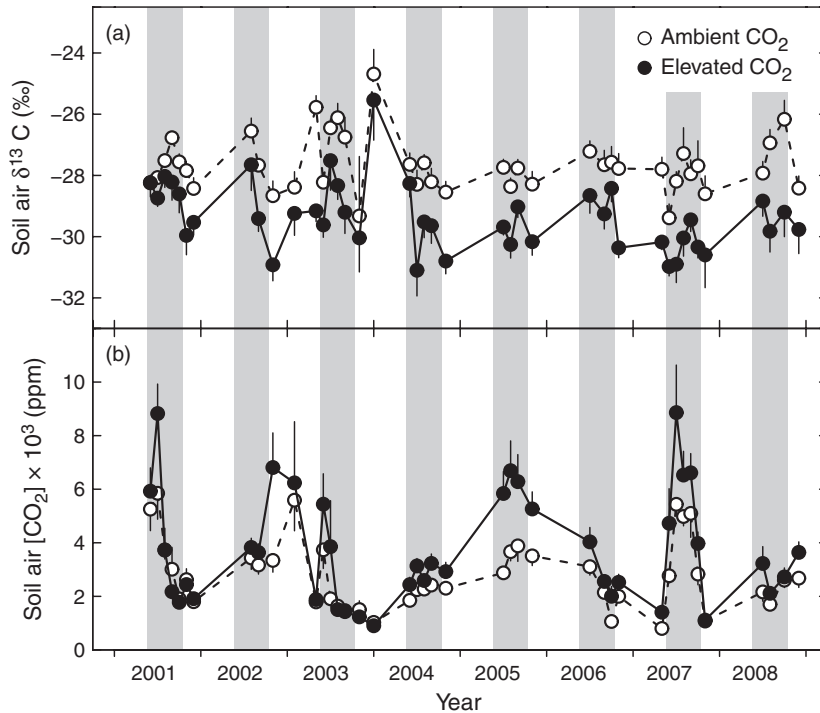


Fig. 7. Interannual and intra-annual variation in the stable carbon isotope signature ($\delta^{13}\text{C}$) (a) and CO_2 concentration (b) in the soil pore space at 3–11 cm depth under CO_2 -enriched and control trees. Means \pm SE, ambient CO_2 $n = 43$ –58, elevated CO_2 $n = 17$ –28. The grey-shaded rectangles indicate the CO_2 -enrichment periods that correspond to the 6-month growing seasons.

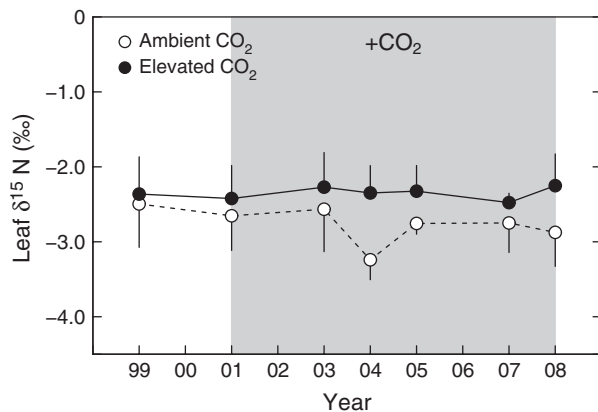


Fig. 8. Nitrogen availability given as $\delta^{15}\text{N}$ of freshly harvested foliage. Symbols indicate composite sample means of the replicated *Fagus sylvatica*, *Quercus petraea* and *Carpinus betulus* trees. Means \pm SE, ambient CO_2 $n = 3$ –5, elevated CO_2 $n = 3$.

production, a signal that also became insignificant towards the end of the experiment (Norby *et al.* 2010). Mediterranean *Quercus ilex* trees growing around two natural CO_2 -springs as well as CO_2 -enriched *Populus tremuloides* clones at the ASPEN FACE site both showed initial growth enhancement that disappeared with time (Hättenschwiler *et al.* 1997; Kubiske *et al.* 2006). Species-specific growth responses became strikingly apparent at the Swiss tree line FACE site where tree-ring increments in the late-successional *Pinus uncinata* remained completely unaffected by elevated CO_2 over 9 years, while growth in early successional *Larix decidua* trees was stimulated (though diminishing with time), depending on summer temperature (Dawes *et al.* 2011, 2013). At the Bangor FACE site, above-ground woody biomass of CO_2 -enriched *Fagus sylvatica* saplings increased only in

monoculture (+22%) but showed no response when saplings were grown in polyculture (Smith *et al.* 2013a). Although there was no sustained growth response in CO_2 -enriched trees at our SCC FACE site, the $\delta^{13}\text{C}$ signal in newly formed tree-rings declined markedly and very rapidly following the start of CO_2 enrichment and quickly returned to pre-treatment levels after the end of the experiment, highlighting the tight coupling between recent photosynthates and wood formation. Yet, the data also illustrate that C ending up in tree-rings has been assimilated over a period of 2–3 years, implying that the current year only contributes a certain percentage to the total bulk signal. The post-treatment return path shown in Fig. 4 depicts this important finding most clearly and suggests that climate correlates of seasonal isotope signals in tree-rings should be roughly halved in strength due to such mixing of photo-assimilates.

ELEVATED CO_2 EFFECTS ON TREE WATER RELATIONS AND SOIL MOISTURE

Many plants reduce stomatal conductance and thus leaf transpiration in response to elevated CO_2 (Medlyn *et al.* 2001; Holtum & Winter 2010). In our study trees, stomatal conductance was slightly diminished by CO_2 enrichment on bright days, with species-specific responses between zero in *Q. petraea* and 22% in *C. betulus* (Keel *et al.* 2007). Similarly, whole-tree transpiration estimated from sap flow measurements was on average 10–15% reduced under elevated CO_2 , resulting in measurable soil water savings (Fig. 5; Leuzinger & Körner 2007; Bader, Hiltbrunner & Körner 2009). These findings are corroborated by other closed-canopy forest FACE experiments and modelling approaches (Warren *et al.* 2011). Water savings on a full year/all weather basis are much

smaller, however. Model simulations based on historical rainfall patterns showed that rainfall distribution by far outweighs the remaining, small (< 3%) effects of CO₂-related soil water savings on runoff in our study forest (Leuzinger & Körner 2010). The main effects of CO₂-driven water savings are below-ground processes discussed below.

BELOW-GROUND RESPONSES TO CO₂ ENRICHMENT

Because of the largely lacking above-ground growth stimulation in our study trees, we assumed greater C investments below-ground. Indeed, proxy data such as higher soil CO₂ concentrations suggested such a response during early experimental years (Steinmann *et al.* 2004; Keel, Siegwolf & Körner 2006). However, we found a 20–30% reduction in fine root biomass and new fine root growth into root-free soil cores under CO₂-enriched trees in year five and six of the study and no significant treatment-related differences in soil cores taken in the following year (Bader, Hiltbrunner & Körner 2009). Similarly, elevated CO₂ failed to stimulate fine root investments in larch and pine trees receiving CO₂ enrichment at the tree line in the Swiss Central Alps and also in a Florida scrub oak system (Brown *et al.* 2007; Handa, Hagedorn & Hättenschwiler 2008). At the Bangor FACE site, CO₂ enrichment merely produced a transient stimulation in fine root biomass of *Fagus sylvatica* saplings and even caused a decline in their coarse root biomass (Smith *et al.* 2013b). Fine root production dominated the CO₂-driven stimulation of NPP over several years of CO₂ enrichment at the Oak Ridge *Liquidambar styraciflua* plantation, particularly at greater soil depth. But this response ceased during the last years of the study and might therefore reflect a transitory response to a step change in CO₂ supply (Norby *et al.* 2010).

The fossil CO₂ used for canopy enrichment was consistently depleted in ¹³C relative to ambient atmospheric CO₂ and thus allowed us to trace the flow of newly assimilated (labelled) C from the crowns to the rhizosphere of the treated trees (Steinmann *et al.* 2004; Keel, Siegwolf & Körner 2006). Only 11 days after the start of CO₂ enrichment, the isotopic signal of the fumigation gas became detectable in soil air (Steinmann *et al.* 2004) and after 4 growing seasons, newly developed leaves consisted of 100%, and new tree-rings of 91% recent C (Keel, Siegwolf & Körner 2006). In fine roots (< 1 mm) formed in ingrowth cores during years five and six of the study, only 51% of the C carried the isotopic signature of the CO₂ released in the canopy, implying long fine root C turnover rates of *c.* 12 years or utilization of old non-structural carbon reserves for root formation (Bader, Hiltbrunner & Körner 2009).

Recent photosynthates were rapidly channelled to below-ground C sinks, as evidenced by strong isotopic signals in soil air and symbiotic fungi (Steinmann *et al.* 2004; Keel, Siegwolf & Körner 2006). Three months after the beginning of CO₂ enrichment, sporocarps of mycorrhizal fungi associated with CO₂-enriched trees already consisted of 62% new C, while saprophytic fungi were devoid of ¹³C-depleted C after 4 years of treatment suggesting decomposition of C

compounds that were formed prior to the start of the experiment (Keel, Siegwolf & Körner 2006).

Given the absence of increases in above- and below-ground growth and litter production, we assumed that the extra C assimilated under elevated CO₂ might be largely respired back to the atmosphere via soil metabolism (Körner *et al.* 2005). Surprisingly, the stronger CO₂ build-up in soil under CO₂-enriched trees during growing seasons did not translate into a sustained increase in soil respiration, most likely due to reduced soil diffusivity resulting from soil water savings (Bader & Körner 2010). This contrasts with other forest FACE studies where soil CO₂ efflux in closed-canopy stands increased by 12–23% (King *et al.* 2004; Jackson *et al.* 2009). Leaching of DIC and DOC was not a major loss pathway for the ‘missing C’ either. The total fluxes of dissolved C were small (*c.* 15 g C m⁻² a⁻¹) and remained unaffected by the CO₂ enrichment (Fig. 6). In the topsoil, CO₂ enrichment increased DIC fluxes by 50%, most likely reflecting the higher partial CO₂ pressure in soil pores. This increase was outbalanced by an unexpected decline in DOC leaching, which we primarily attribute to reduced solubilization of soil organic matter (SOM) in the mineral soil through acidification associated with increased soil CO₂ concentrations. Topsoil acidification was further enhanced by higher NO₃⁻-N leaching as two protons are released per mole of nitrate leached, thereby suppressing DOC leaching (Evans *et al.* 2008). At 15 cm depth, both DOC and DIC leaching remained unaffected by CO₂ enrichment, thus ruling out this ‘leak’ pathway for the additionally assimilated C, perhaps because this forest grows on shallow soil over calcareous rock and rock debris. At the Aspen and Duke FACE sites, DIC leaching increased considerably under elevated CO₂ (Karberg *et al.* 2005; Jackson *et al.* 2009).

There is a growing awareness that CO₂ fertilization effects will be smaller than anticipated (Leuzinger *et al.* 2011) and will certainly not be uniform across landscapes but rather follow availability patterns of colimiting growth resources such as water and nutrients (Oren *et al.* 2001; Körner 2006; McCarthy *et al.* 2010). Moreover, potential CO₂ fertilization effects on tree growth might be offset over time by progressive N limitation in soils resulting from N sequestration in long-lived biomass or SOM pools or by reduced longevity (Luo *et al.* 2004; Norby *et al.* 2010; Bugmann & Bigler 2011). For instance, after 9 years of CO₂ enrichment at the Oak Ridge site, progressive N limitation caused a decline of the strong NPP response that was previously dominated by fine root production (Norby *et al.* 2010). At our site however, the lacking growth responses to CO₂ enrichment cannot be attributed to N deficiency since this region receives 20–25 kg N ha⁻¹ a⁻¹ wet nitrogen deposition, which is close to the upper threshold of critical loads for this type of ecosystem (Thimonier *et al.* 2010). Moreover, CO₂-induced increases in soil moisture and nitrate availability resulted in enhanced nitrate leaching at this already N-rich site. Also, the increase in the natural abundance of ¹⁵N in nitrate captured by resin bags suggests accelerated net nitrification under elevated CO₂ (Schleppi *et al.* 2012). The temporal δ¹⁵N dynamics derived from archived leaf samples of our tall study trees remained remarkably stable under

elevated CO₂ suggesting unaltered and ample N supply even during years when lower δ¹⁵N values of control foliage indicated declining N availability under ambient conditions (Fig. 8). This was most obvious in 2004, probably representing a carry-over effect from the preceding centennial drought year 2003 that most likely had a greater impact on soil N cycling processes under control trees compared to CO₂-enriched trees simply because of the soil water savings observed under elevated CO₂ (Leuzinger *et al.* 2005). Reductions in fine root biomass may be associated with diminished nitrate consumption under elevated CO₂, however, given the largely unaltered leaf N concentrations, accelerated SOM decomposition offers a more likely explanation for the accumulation of nitrate in the soil solution under high CO₂. This assumption is supported by a 14% increase in soil microbial biomass under CO₂-enriched trees (Bader & Körner 2010). Based on our and other FACE data, we postulate two concurrent causes underlying the observed stimulation of soil microbes and soil nitrate accretion. CO₂-induced soil water savings may have stimulated soil microbial biomass and nitrification rates, which have been shown to increase with increasing soil moisture (Stark & Firestone 1995). In addition to this water-driven effect, CO₂-enriched trees can fuel microbial activity in their rhizosphere through enhanced root exudation as has been demonstrated for tropical woody species in large model communities (Körner & Arnone 1992) and more recently for *Pinus taeda* at the Duke FACE site (Phillips, Finzi & Bernhardt 2011). There, this priming effect caused faster rates of SOM mineralization and nitrification, which prevented (at least for the period considered) progressive nitrogen limitation in the inherently N-poor forest soil (Drake *et al.* 2011). Increases in soil solution nitrate at our site may have been driven by such a priming effect stimulating microbial activity and thus N mineralization. The decrease in DON that paralleled the increase in soil nitrate supports this hypothesis.

In conclusion, our data imply that tree growth in this late-successional temperate forest is not limited by current atmospheric CO₂ concentrations, suggesting that these types of stands are unlikely to grow faster in a future high-CO₂ world. Whether or not tree growth will be stimulated, such experiments are principally unable to infer long-term changes in ecosystem C pools (C sequestration, Körner 2006), since C storage in biomass is in essence a demographic phenomenon (Bugmann & Bigler 2011), and soil C pools are responding too slowly to allow separation of transient from steady-state responses. It remains unresolved which resources become rate-limiting under humid, C and N saturated conditions, but phosphate and key cations are likely candidates. Our findings point towards enhanced DIC and nitrate leaching from such forest soils with rising atmospheric CO₂ levels. As a result, soil acidity and mineral weathering may intensify in a CO₂-rich future impacting on soil biota and nutrient cycling, while at the same time, nitrate contamination of groundwater is likely to become an increasing problem, especially in already predisposed regions.

Overall, we conclude that indirect CO₂ effects inducing changes in the forest N-cycle and soil water regime are far more

important than first-order CO₂ effects on growth in this system. With regard to future climate predictions, our findings and those from other FACE studies challenge the assumption of a sustained and universal CO₂ fertilization effect on tree growth that most climate–carbon cycle models critically rely on.

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